Cryptosporidium spp. in Cattle in a Jamaican Watershed

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Background

- Cryptosporidium is a protozoan parasite associated with gastrointestinal illness in humans and a wide range of vertebrate animals.
- Cattle, particularly calves, are widely recognized as major reservoirs of zoonotic *Cryptosporidium spp*.
- In the Caribbean region, molecular characterization of *Cryptosporidium spp.* in cattle is limited and their zoonotic potential is poorly understood.

Objectives

 To perform an exploratory investigation on the presence and molecular characterization of *Cryptosporidium spp.* in dairy and beef cattle in a Jamaican watershed and determine whether cattle are potential zoonotic reservoirs for human infection on the island.

Methods

- A total of 119 fecal specimens were collected from 60 dairy and 59 beef cattle from 10 farms in the watershed (Fig 1).
- Approximate age and absence/presence of diarrhea was recorded from each specimen.
- Phosphate-buffered saline (PBS)-ether sedimentation was performed to concentrate potential *Cryptosporidium* ooysts from individual specimens.
- Initial screening for *Cryptosporidium* was performed by modified acid-fast (MAF) staining microscopy, followed by conventional and nested PCR amplification of a polymorphic locus on the 18S rRNA gene and sequencing.
- A phylogenetic tree was constructed using neighbourjoining in MEGA 11, based on Kimura's two-parameter model with bootstrap values obtained from 1000 replicates.
- Further subtyping analysis of PCR-positive isolates was performed by *gp60* nested PCR and sequencing.
- The *gp60* subtype was identified based on family designation and the number of trinucleotide repeats (TCA or TCG) in the sequence.





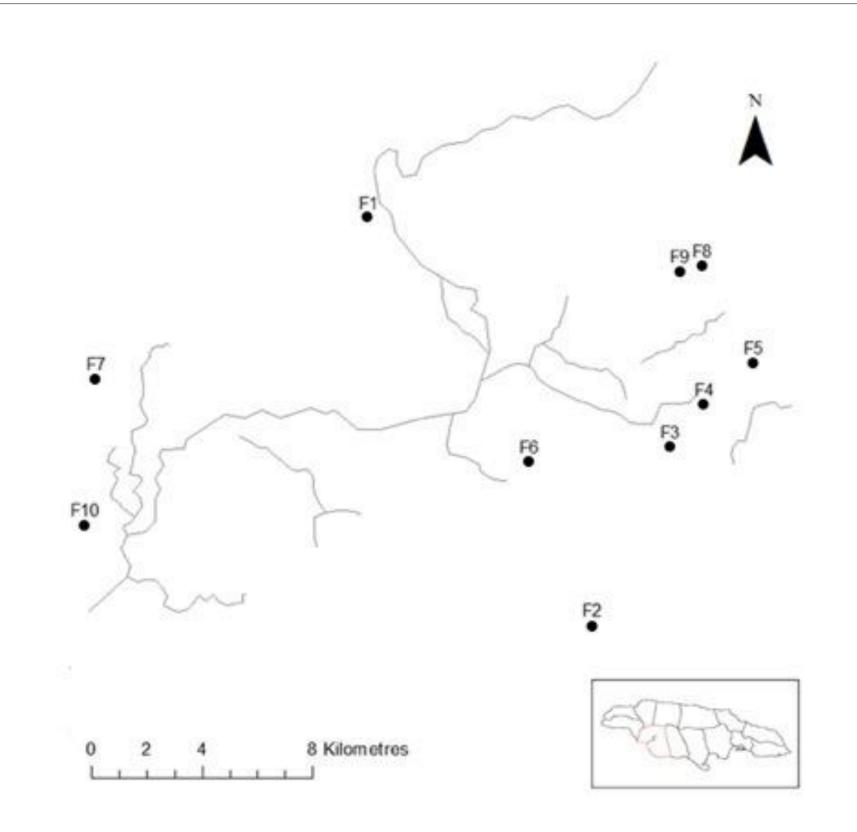


Fig 1. Map illustrating sampling sites in agricultural watershed in St. Elizabeth, Jamaica

Fig 2. Representative images of *Cryptosporidium* oocysts detected from cattle sample 56DaCw by PBSether sedimentation and MAF staining. The Ward's® Chemistry Acid-Fast Stain Kit was used to stain oocysts, as per the CDC modified acid-fast staining procedure. Oocysts have a well-formed wall with diameter of approximately 4-5 µm. 100X with immersion oil.

Table 1 Cryptosporidium spp. identified from cattle by PCR and sequence analysis of the 18S rRNA and gp60 loci

	anarysis						
				18S rRNA		Gp60	
	Farm ID	Sample ID	GenBank accession no.	Species (% identity)	GenBank accession no.	Species (subtype ID)	Reference
	F3	105Da ^{b,c}		C. hominis (82 %)	MN836824; MK982514	<i>C. hominis</i> (IbA9G2)	Razakandrain ibe et al. 2018
	F4	8Be ^{a,c}	OK361786	C. parvum (90 %)	MW947436		
		13Da 40Da ^c					
		46Be					
	F7	53Da ^{a,c}	OK325584	C. parvum (99 %)	MF074701; AB513881		
		56Da ^{b,c}					
	F9	34Da ^{b,c}	OK310618	C. parvum (99 %)	MF074701; AB513881		
		78Da ^b					
	F10	64Be ^c					
		d calf ≤ 6 months lf ≤ 2 years > 6 mor ymptoms	nths				
				96 - OK310618 Crypto	osporidium parvum 34L	Da ★	
			90		225584 1		
				0x.	325584 Cryptosporidium	i parvum 35Da 🗙	
			52	MF074701 Cryptosporidiu	m parvum		
				OK361786 Cryptos	poridium parvum 8Be 🕇	•	
			72 53 M(3516758 Cryptosporidium ho	minie		
				1910790 Cryptosportation not			
			90 AY741.	305 Cryptosporidium bovis			
			64 EU41	0344 Cryptosporidium ryanae	,		
			AF11.	2575 Cryptosporidium felis			
			86 AF1125	76 Cryptosporidium canis			
			- AY642591 Cry	ptosporidium muris			
			100 AB777194 Crypto				
			100 - AB777194 Crypic	sportatum anaersoni			
				AF0	26388 Eimeria tenella		
		⊢					
		0.01					
with the Eim	enetic tree of e <i>eria tenella</i> ou the present st	tgroup speci	e. Each speci	es is marked w	ith an ident	tifying acces	ssion numbe

e tree was rooted er. Sequences 0.01 substitutions

- from dairy cattle.
- et al., 2018).

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Toronto Metropolitan University

Results

• Of the 119 cattle fecal specimens, 10 (8 %) were confirmed positive for *Cryptosporidium* infection by PCR.

Of the 10 PCR-positive samples, 7 were from cattle presenting symptoms of diarrhea at the time of sample collection; 5 were from calves \leq 2 years of age, including two pre-weaned calves ≤ 6 months (**Table 1**); and 7 were

Sequence analysis of the 18S rRNA gene locus, identified *C. parvum* from one beef calf (no. Be8) and two dairy calves (nos. 34Da and 53Da). Phylogenetic analysis confirmed that *C. parvum* isolates, 34Da and 53Da, share a 96 % nucleotide sequence identity (Fig 3).

• Further sequence analysis of the *gp60* gene, identified the *C. hominis* IbA9G2 subtype in dairy calf no.105Da, previously reported in an impaired calf (Razakandrainibe

Conclusions

This study provides first insight into *Cryptosporidium spp*. infecting Jamaican dairy and beef cattle. Preliminary data suggest cattle, particularly impaired dairy calves, may be important reservoirs of zoonotic *Cryptosporidium spp*. However, the low prevalence of characterized isolates cannot conclude any correlation.

Additional studies, with a larger number of PCR-positive isolates from calves are necessary to provide critical information about geographically distinct, clonal genotypes of Cryptosporidium spp., which have potential to cause zoonosis in the country.

With meaningful interpretations of Cryptosporidium population structures, useful databases can be built through analysis of well-planned sets of human and environmental samples.

Acknowledgements