MOLECULAR EPIDEMIOLOGY OF AMOEBIC GILL DISEASE CAUSING NEOPARAMOEBA PERURANS IN ATLANTIC SALMON AQUACULTURE

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Introduction	Aim	Methodology		
 Amoebic Gill Disease (AGD) → Caused by Neoparamoeba perurans Symptoms include: Pale lesions → raised white mucoid patches on the gill → Lethargy → Lack of appetite → Respiratory distress → Death AGD affects a variety of marine fish and cleaner fish not only Atlantic salmon Downstream effects of such high levels of AGD infection and resultant suppressed immune function 	Using the Llewellyn lab's draft genome of <i>N. perurans</i> , develop a amp-seq SNP panel to genotype the parasite and enable near real-time information to inform disease control management plans including when are where to fallow for Atlantic salmon aquaculture.	 Gill swab collection DNA extraction and Quantification (DNAeasy & Qubit) RT-qPCR – confirm the presence (ng/μl) & aid selection of positive samples for sequencing Multiplex PCR 96 experimental primers, mapped against <i>N. perurans</i> draft genome in the Llewellyn lab Success in amplifying up to 48 primers at a 	 Sequencing Nanopore MinION Demultiplexed and trimmed IGV used to view primer pile up plots Aligned using MUSCLE (Jalview) Molecular Epidemiology Phylogenies built-in MEGA11 	

can lead to greater susceptibility to other marine pests, such as salmon lice

University of Glasgow

 Predicted to cost the UK salmon farming industry £124 million in 2023 in losses

UNDERSTAND EPIDEMIOLOGY

 Ascertaining the transmission and evolutionary pathways of this amoeba is of great importance to prevention efforts.

Current

uploaded data

Management zone is incorrect

Inputted

data



Ireland

2019 sample

2022 sample

2023 sample

Green Box

Drange Highlight

ellow Highlight

Pink Highlight

Symbol	Location/Year	bootstrap value		Irish_B2	
Blue Boy	Scotland	0.946	▶ 0.77		



Figure 1: Bar graphs visualizing the number of reads per primer loci

Results

- Primers cleary amplify *N. perurans* genetic material.
- Cyclical reintroduction and reinfection of the same amoeba strains and genetic material year over year may occur.

 Phylogenetic analysis indicates significant genetic mixing and reinfection year over year in the Scottish and Irish marine environment



(labeled 1-96) as recorded via Nanopore MinION sequencing for each gill swab sample sequenced

Figure 2: Evolutionary trees of *N. perurans* using a bootstrapped Neighbor-Joining method (A) and a Maximum Likelihood (ML) method (B) where bootstrap values and branch lengths are displayed respectively. Key top left.

Further work

Take Home Message(s)

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- Further research on primer design to discover why many of the primers aren't amplifying, and which ones are working sporadically
- Test GLST AmpSeq approach using 202 experimental primers on tissue and gill swab samples from different years and sites across 3 countries (Scotland, Ireland, & Norway) to understand spread and genomic variation over time.
- Sequencing using Illumina \rightarrow more suited for small fragments of DNA
- Development of a web-based data visualisation tool to track genotypes, spread over time, and new more virulent lineages of *N. perurans* for Atlantic salmon farming sites (potential to be used worldwide) can enable targeted and timely approaches to AGD outbreaks.
- Phylogenetic analysis of trees created from useable consensus sequences indicate reinfection and significant genetic mixing and similarity year over year in the Scotland and Ireland marine environment
 - Which would **negatively affect the efficacy of prophylactic treatment strategies for AGD** and recommend practices centered around freshwater fallowing and general cage hygiene
- Appears to be strong evidence that AGD treatment in aquaculture should remain focused on cage hygiene and freshwater fallowing as needed.
- Information obtained from the GLST AmpSeq panel on *N. perurans* dispersal can be relayed back to the individual farm managers of affected sites (and potentially other site managers nearby) in near real-time to allow appropriate decisions to be made i.e. treatment protocols (freshwater bathing), emergency harvest, and fallowing.