

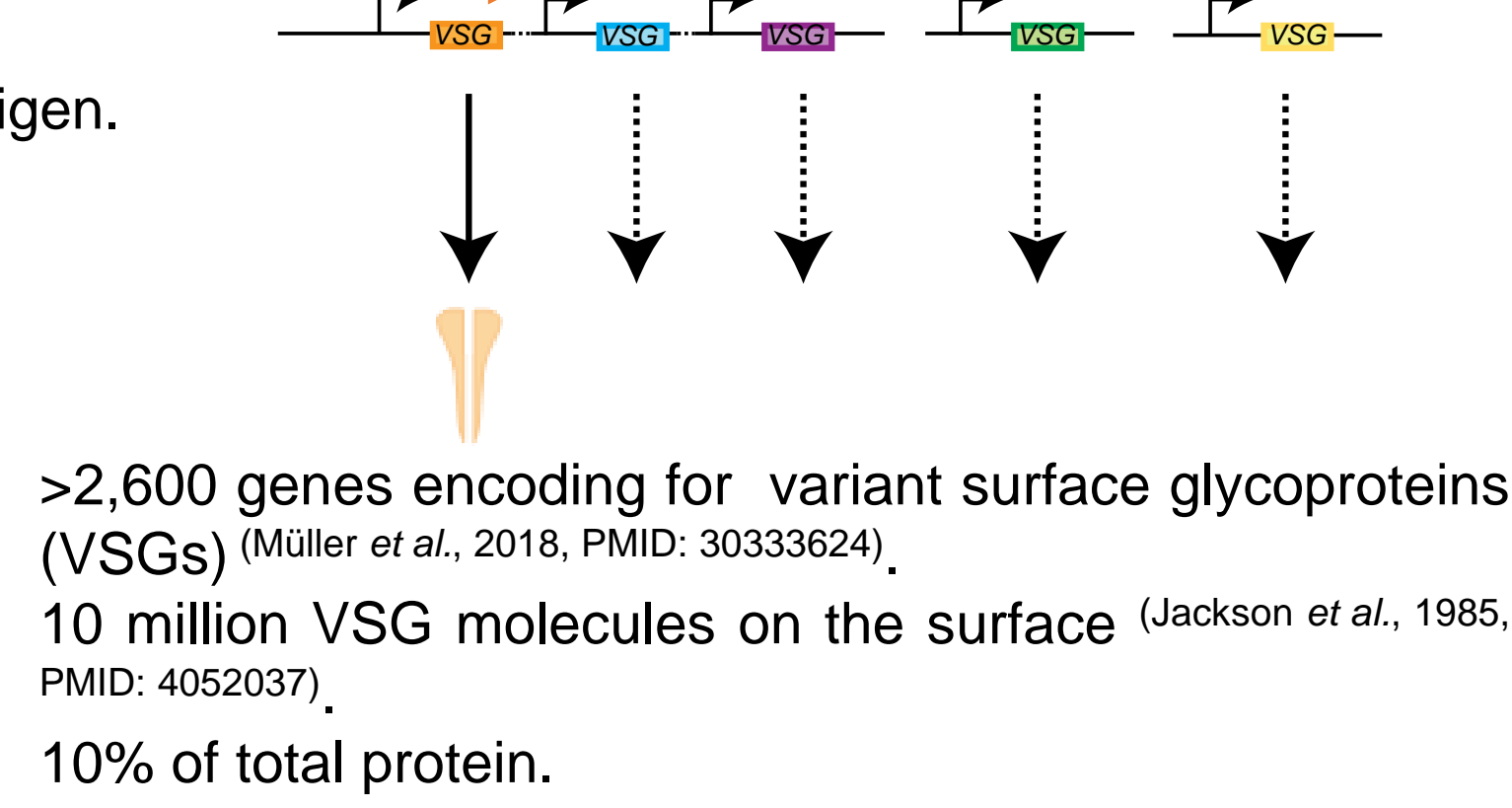
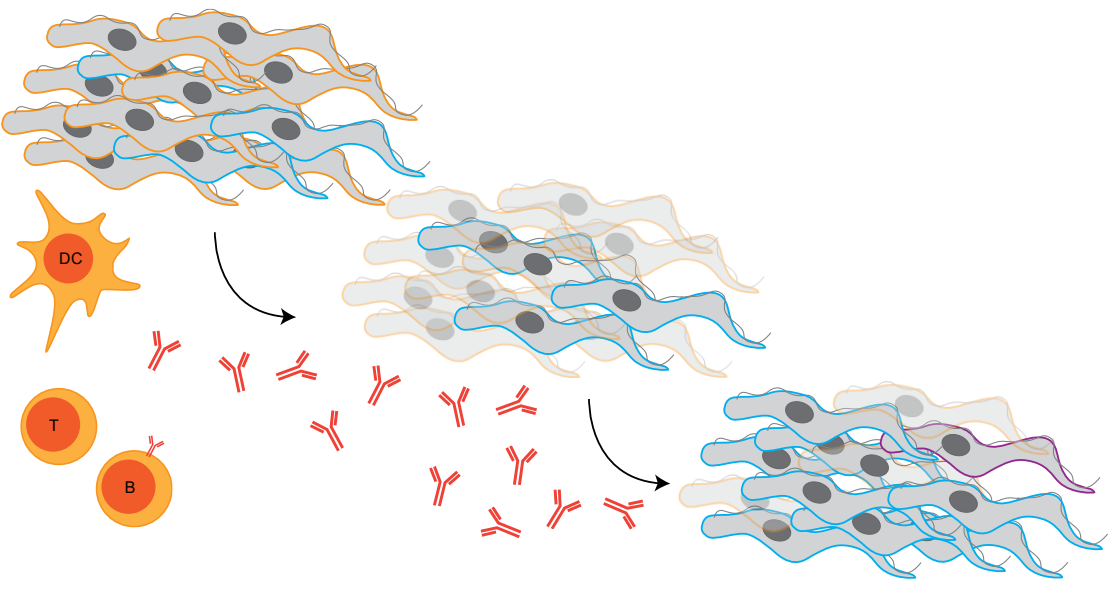
Novel components of the expression-site body and surrounding splicing bodies discovered by TurboID proximity labelling in African Trypanosomes.

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1. Antigenic variation in African Trypanosomes and the role of VEX in monogenic expression

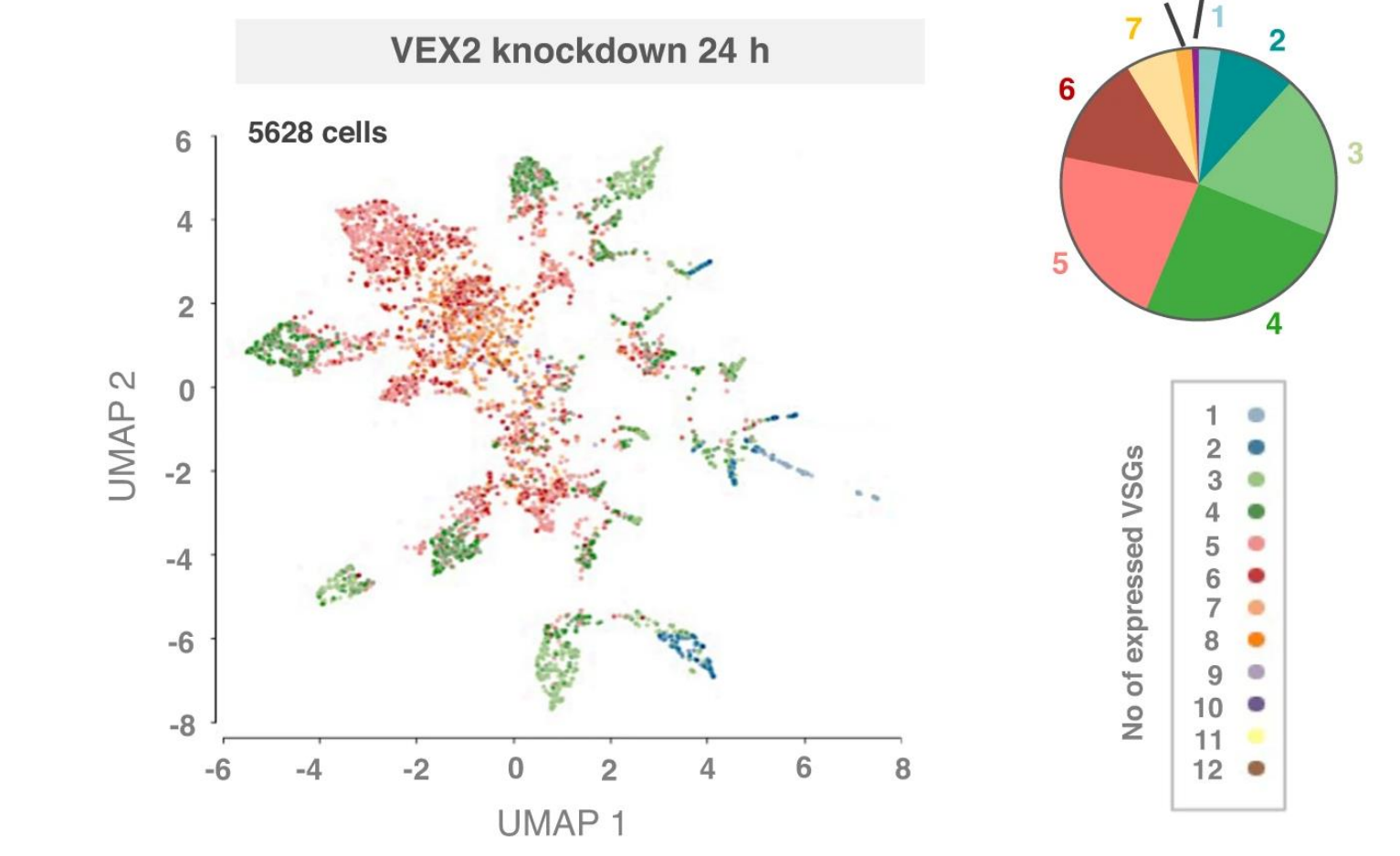
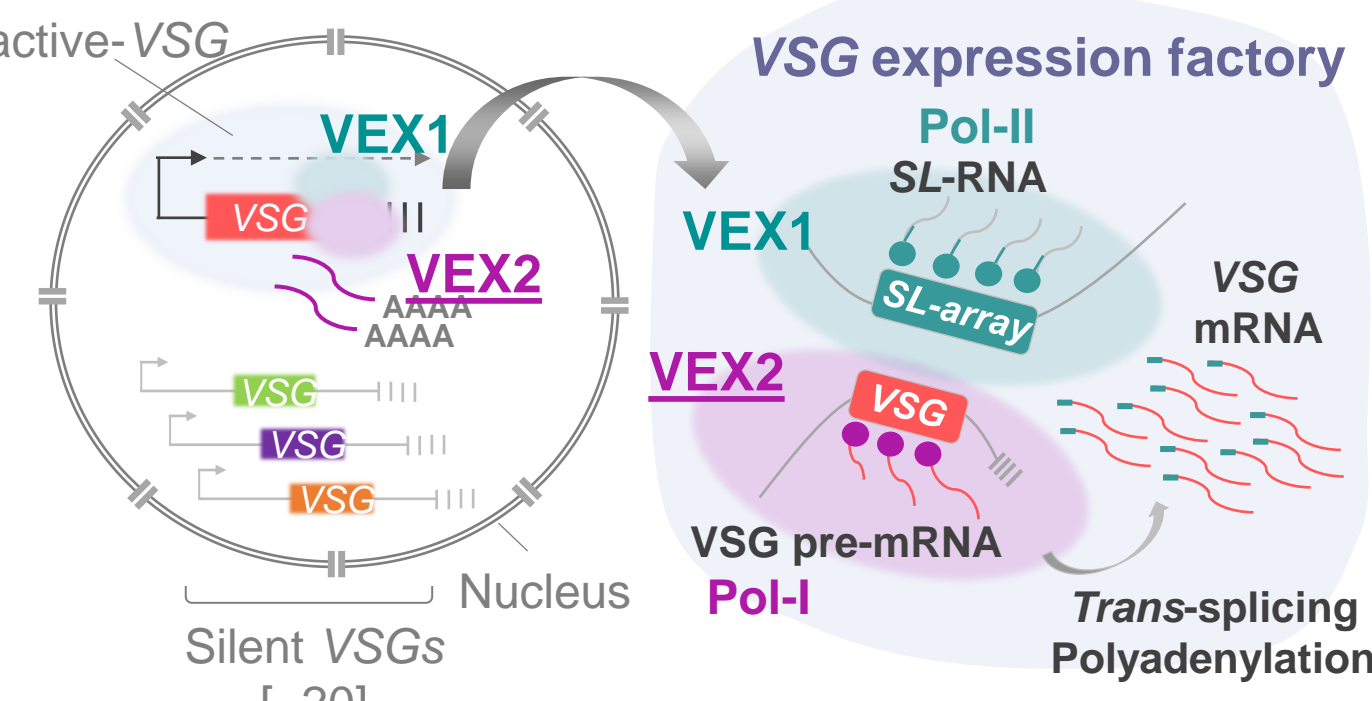
Successful antigenic variation depends on:
1. Monogenic expression of a surface-exposed-antigen.
2. Switching of the antigen coat.



VEX2 KD results in derepression of 'silent' VSGs

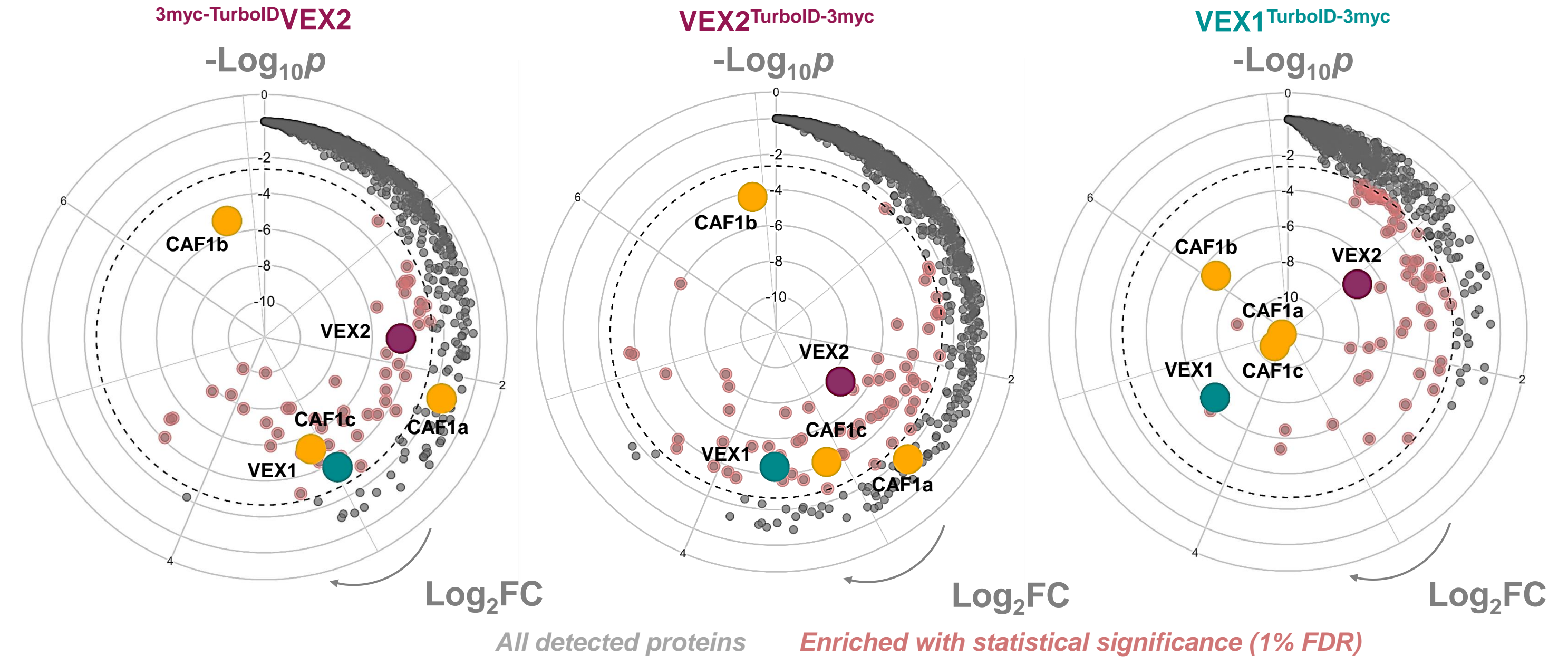
VEX2 enforces monogenic expression in *T. brucei* parasites. Its depletion leads to simultaneous expression of multiple VSG genes (Faria *et al.*, 2019, PMID: 31289266, Faria *et al.*, 2023, PMID: 38081826).

VEX complex bridges the ESB and SLAB
VEX1 and VEX2 form a dynamic complex that brings together the active-VSG expression-site and the SL-array (Faria *et al.*, 2021, PMID: 33432154). The latter is necessary for trans-splicing and mRNA maturation.



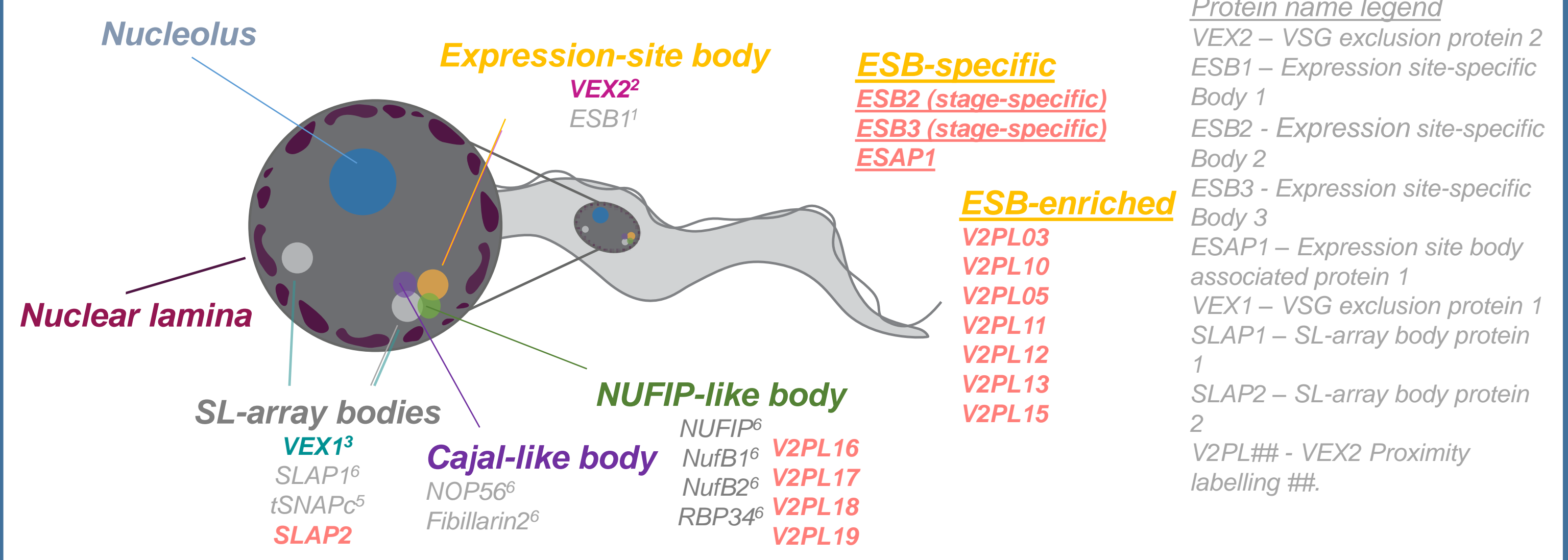
2. Identification of the spatial interactome of the 'VSG expression factory' using proximity labelling

Two biotin ligases, TurboID (Doerr, 2018, PMID: 30275580) and UltraID (Zhao *et al.*, 2021, PMID: 35788163), were tested. TurboID outperformed UltraID (data not shown).



Proximity labelling data shows an enrichment of previously identified VEX2 interactors, VEX1 and the CAF-1 subunits. Additionally, the dataset revealed a range of previously unidentified hypothetical proteins that form part of the VSG expression factory.

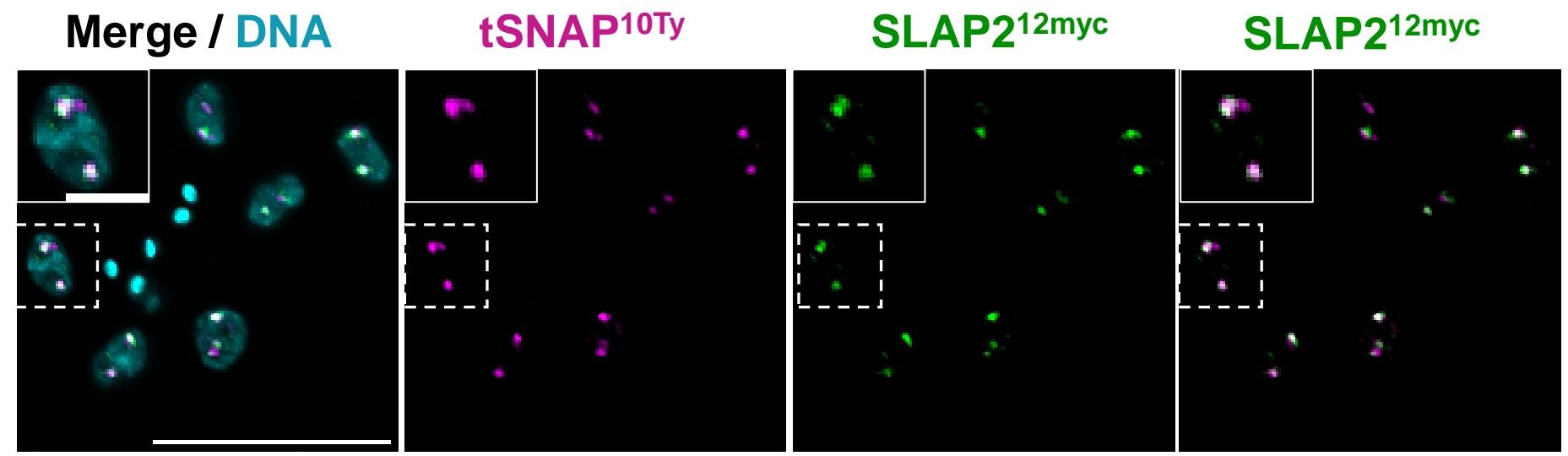
VEX2 proximity labelling reveals known proximal proteins and a range of hypothetical proteins



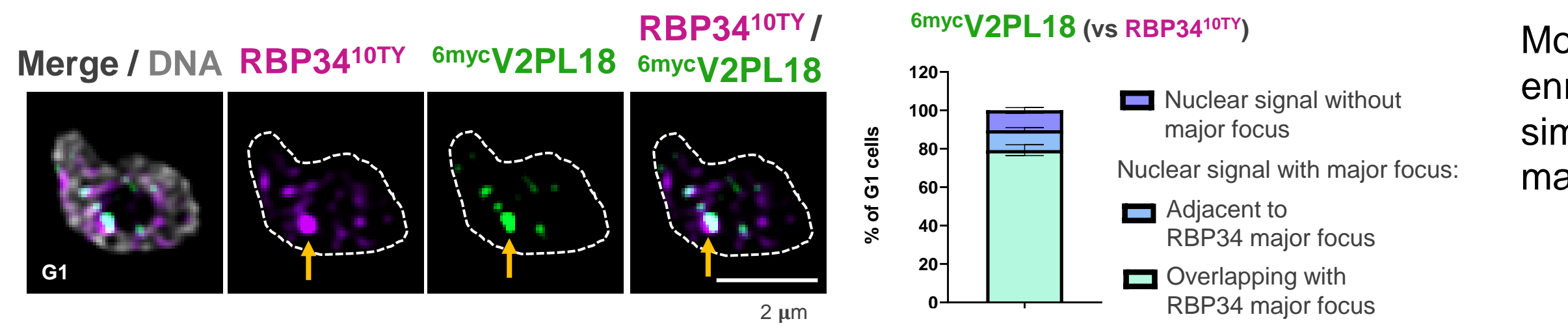
3. Novel SLAB and NUFIP components

SLAP2 localises specifically to the SLAB

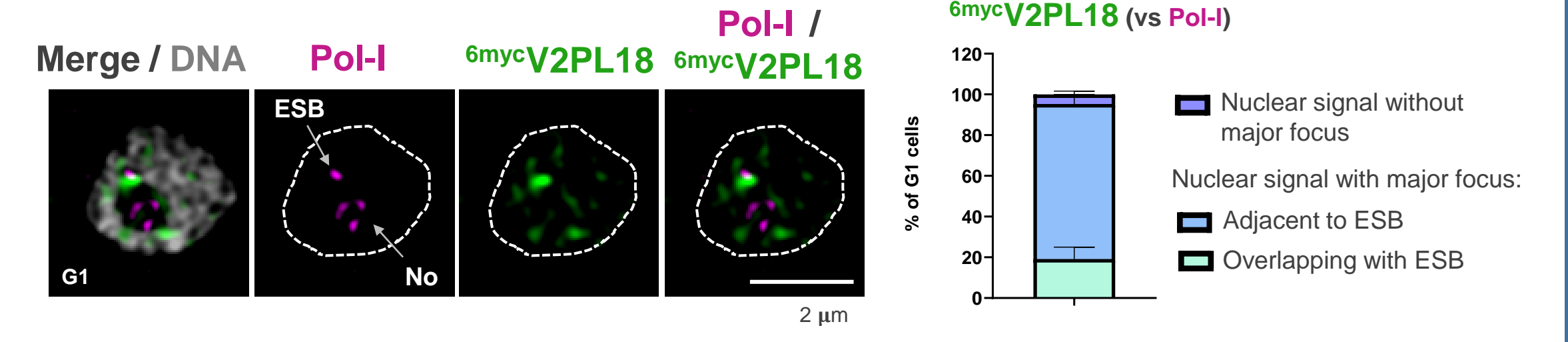
Proximity labelling also revealed a range of components found to localise to other splicing bodies associated with VSG expression. One such new component, SLAP2, overlaps with tSNAP.



Four proteins (V2PL16-V2PL19) were found to colocalise with RBP34, a marker of the NUFIP 'body'.

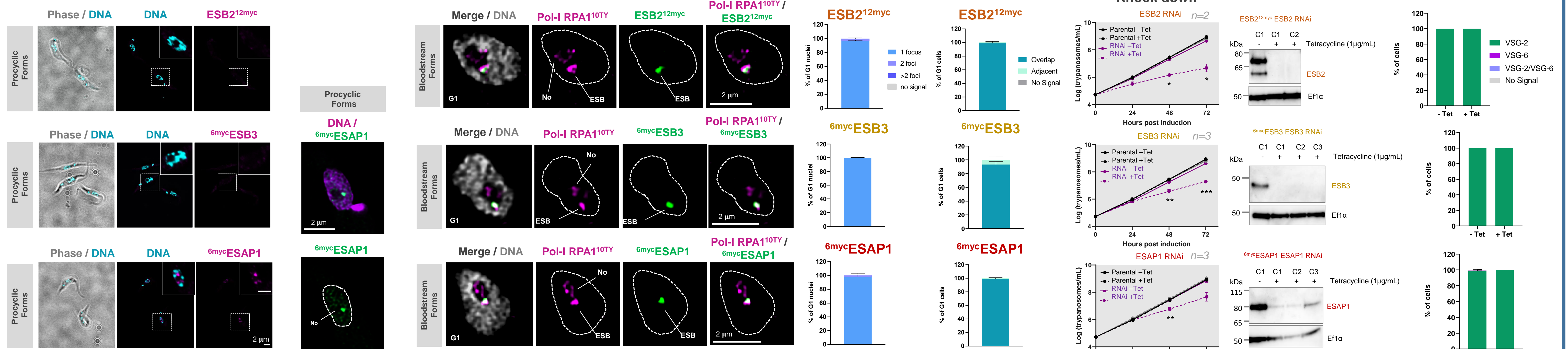


Moreover, these proteins were enriched adjacent to the ESB, similarly to RBP34 (not shown), marked by Pol-I.

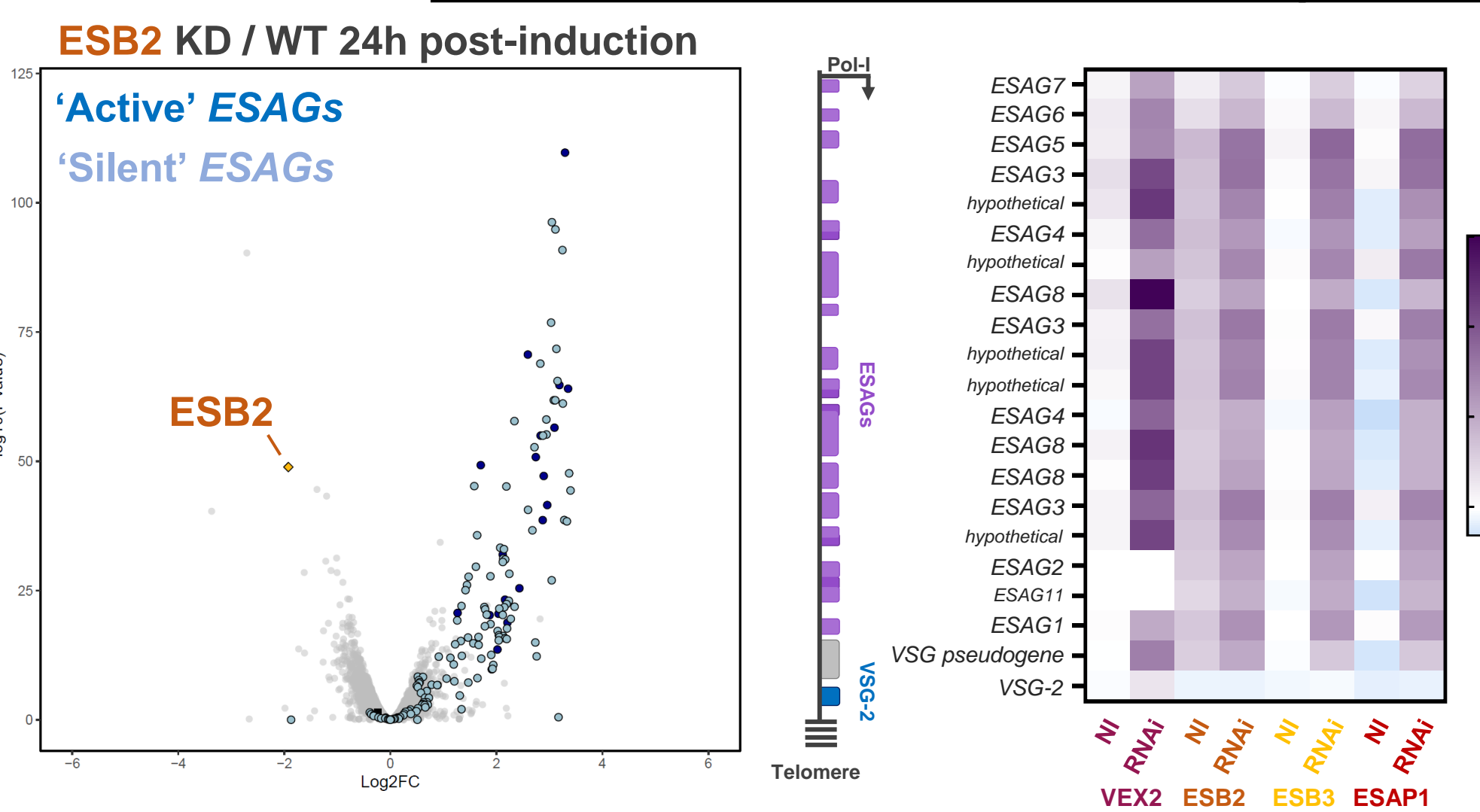


4. Novel ESB components: ESB2, ESB3 and ESAP1

ESB2, ESB3 and ESAP1, identified by VEX2 proximity labelling, were all found to specifically localise to the ESB, marked by Pol-I expression, in bloodstream forms. RNAi knock down of each protein resulted in significant fitness loss.

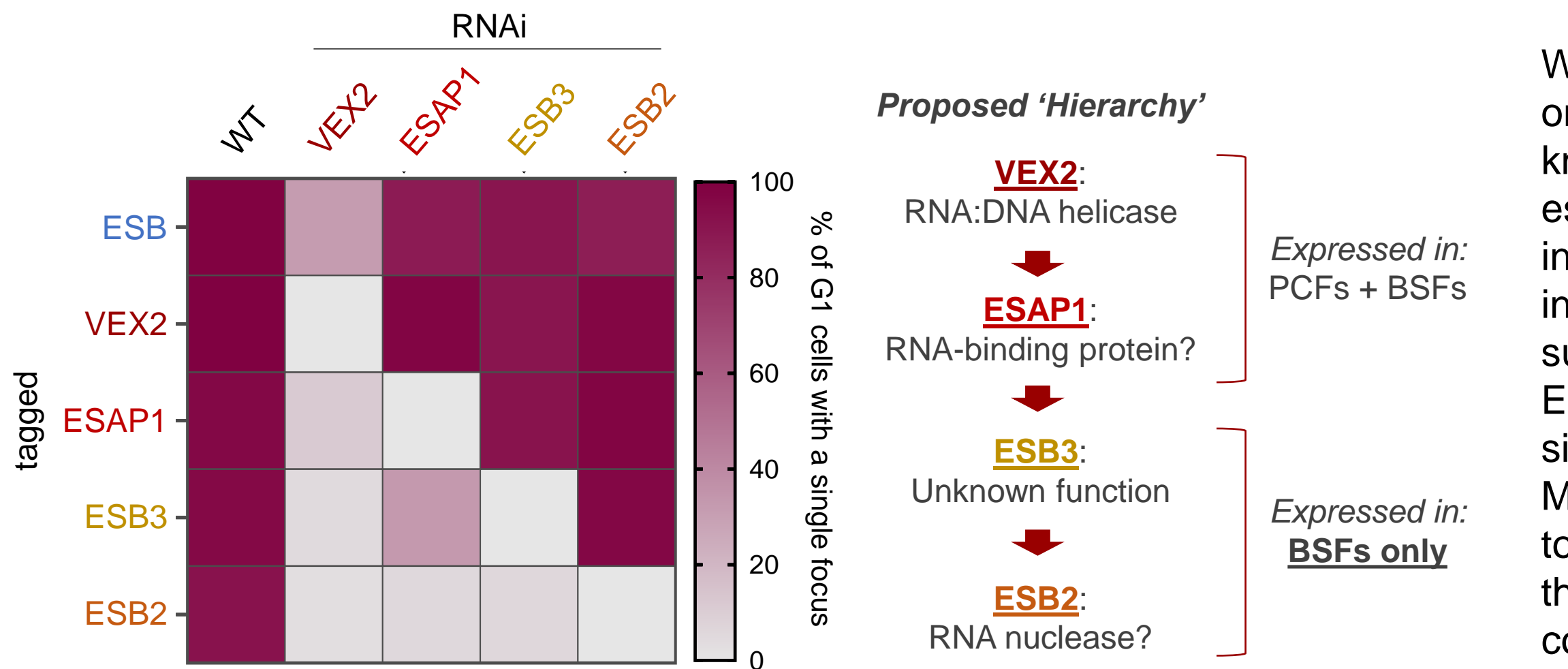


ESB2, ESB3 and ESAP1 KDs lead to specific upregulation of ESAGs



ESB2, ESB3 and ESAP1 knock down each result in the upregulation of both active and 'silent' ESAG genes, but do not significantly affect either active or 'silent' VSG gene expression despite sharing the same promoter(s) per polycistron. Patterns of upregulation per ESAG are similar between the knockdowns of ESB2, ESB3 and ESAP1, but dissimilar to the VEX2 knock down pattern.

'Co-dependencies' between novel ESB factors



We hypothesise that the effect on ESAGs seen after VEX2 knock down are due to the essentiality of VEX2 to the integrity of the ESB, an indirect effect. This is supported by the loss of focal ESB2, ESB3 and ESAP1 signal after VEX2 knock down. Moreover, a hierarchy seems to exist within the group of three newly discovered ESB components.

5. Summary & Future directions

Uncovering molecular mechanisms behind ESB2-mediated ESAG expression regulation.

- Is the ESB2-KD phenotype dependent on the nuclease activity?
- Does ESB2 bind (ESAG) mRNA?
- Does ESB2 affect transcription, splicing and/or degradation of ESAG transcripts?

References

1. López-Escobar *et al.*, 2022, PMID: 35879525. 2. Faria *et al.*, 2019, PMID: 31289266. 3. Glover *et al.*, 2016, PMID: 27226299. 4. Das *et al.*, 2005, PMID: 16055739. 5. Budzak *et al.*, 2022, PMID: 35013170. 6. Jehi *et al.*, 2014, PMID: 24810301.