"Multi-omic approaches reveal a dynamic crosstalk between plasma cells and *Cx3cr1*+ microglia in the brain during chronic *Trypanosoma brucei* infection"

<u>Juan F. Quintana^{1,2}</u>, Praveena Chandrasegaran^{1,2}, Matthew Sinton^{1,2}, Rhiannon Heslop^{1,2}, Emma Briggs⁴, Thomas Otto^{1,3}, Neil Mabbott⁵, Annette MacLeod^{1,2}

¹Wellcome Centre for Integrative Parasitology (WCIP). Institute of Biodiversity, Animal Health, and Comparative Medicine. University of Glasgow, Glasgow, UK. ²Institute of Biodiversity, Animal Health and Comparative Medicine (IBAHCM). University of Glasgow, Glasgow UK.

^{1,3} Institute of Infection, Immunity & Inflammation. University of Glasgow, Glasgow, UK.

⁴ Institute for Immunology and Infection Research, School of Biological Sciences, University of Edinburgh, Edinburgh, UK

⁵The Roslin Institute and Royal (Dick) School of Veterinary Sciences, University of Edinburgh, Edinburgh, UK.

Email: juan.quintana@glasgow.ac.uk

Chronic infections with the parasite Trypanosoma brucei, the causative agent of Human African trypanosomiasis, lead to severe neuroinflammation and death if left untreated. However, a detailed understanding of the cellular and molecular interactions that mediate this severe pathology is lacking. Using single cell and spatial transcriptomics, we have identified for the first time, a unique population of CD138⁺ plasma cells in the brain ventricles of infected animals compared to naïve controls. These plasma cells express a robust innate-like, regulatory transcriptional profile, characterised by the expression of pathogen-sensing molecules (TIr4), antiinflammatory cytokines (II10) and pro-survival receptor molecules such as Tnfrsf17 (B cell maturation antigen, BCMA). Additionally, we detected a subpopulation of Cx3cr1⁺ microglia that express a wide range of factors associated with B cell recruitment and survival, such as Cxcl12 and Tnfsf13b (B cell activating factor). Interestingly, Cx3cr1⁺ microglia are the only cells in our dataset expressing both *II10ra* and *II10rb*, suggesting that they are primed to respond to IL-10. Further in vitro studies demonstrated that these regulatory, innate-like plasma cells can stimulate microglia polarisation towards an anti-inflammatory state via IL-10 signalling. We propose a model in which unresolved brain infections induce the activation of $Cx3cr1^+$ microglia, leading to the recruitment and survival of plasma cells mediated by CXCL12 and BAFF-BCAM signalling, respectively. In turn, these regulatory plasma cells alleviate inflammation by dampening *Cx3cr1*⁺ activation *via* IL-10 signalling, limiting pathology. This work provides novel insights into the mechanisms of B cell-stromal interactions in the brain during infection.